

Freeform Search

10097383

Database:

US Pre-Grant Publication Full-Text Database
 US Patents Full-Text Database
 US OCR Full-Text Database
 EPO Abstracts Database
 JPO Abstracts Database
 Derwent World Patents Index
 IBM Technical Disclosure Bulletins

Term:

l3 and RNaseH

Display:

10

Documents in Display Format:

-

Starting with Number

1

Generate:

☐

Hit List

☒

Hit Count

☐

Side by Side

☐

Image

Search

Clear

Interrupt

Search History

DATE: Thursday, January 06, 2005 [Printable Copy](#) [Create Case](#)

<u>Set</u> <u>Name</u>	<u>Query</u>	<u>Hit</u> <u>Count</u>	<u>Set</u> <u>Name</u> result set
side by side			
	DB=USPT,EPAB,JPAB,DWPI; PLUR=YES; OP=ADJ		
<u>L7</u>	l3 and RNaseH	4	<u>L7</u>
<u>L6</u>	l4 and reverse transcriptase\$1	0	<u>L6</u>
<u>L5</u>	l3 and (reverse transcriptase\$1 near5 (lack\$1 or devoid\$3) near5 RNaseH)	0	<u>L5</u>
<u>L4</u>	L3 and (reverse transcriptase near5 lack\$1 near5 RNaseH)	0	<u>L4</u>
<u>L3</u>	(primer\$1 or oligonucleotide\$1) same (brideg\$3 sequence\$1 or spac\$3 sequence\$1)	239	<u>L3</u>
<u>L2</u>	L1 and (reverse transcriptase\$1 near5 lack\$1 near5 RNaseH)	1	<u>L2</u>
<u>L1</u>	(primer\$1 or oligonucleotide\$1)same (brideg\$3 or spac\$3)	5030	<u>L1</u>

END OF SEARCH HISTORY

10077383

> s (primer# or oligonucleotide#) (P) (bridge### sequence# or spac### sequence#)
 L1 291 (PRIMER# OR OLIGONUCLEOTIDE#) (P) (BRIDGE### SEQUENCE# OR SPAC##
 # SEQUENCE#)

=> s l1 and reverse transcriptase#(10a)lack#(10a)RNaseH
 L2 0 L1 AND REVERSE TRANSCRIPTASE#(10A) LACK#(10A) RNASEH

=> s l1 and (reverse transcriptase#(10a)lack#(10a)RNaseH)
 L3 0 L1 AND (REVERSE TRANSCRIPTASE#(10A) LACK#(10A) RNASEH)

=> s l1 and reverse transcriptase#
 L4 8 L1 AND REVERSE TRANSCRIPTASE#

=> s l4 and RNaseH
 L5 1 L4 AND RNASEH

=> d l5 bib ab kwic

L5 ANSWER 1 OF 1 CAPLUS COPYRIGHT 2005 ACS on STN
 AN 2002:946303 CAPLUS
 DN 138:1057
 TI Nucleic acid amplification utilizing intermediate duplexes
 IN Haydock, Paul V.; U'Ren, Jack
 PA Saigene Corporation, USA
 SO PCT Int. Appl., 69 pp.
 CODEN: PIXXD2
 DT Patent
 LA English
 FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	WO 2002098895	A1	20021212	WO 2002-US18229	20020607
	W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZM, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM				
	RW: GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR, BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG				
	US 2003050444	A1	20030313	US 2002-77383	20020215
	EP 1404697	A1	20040407	EP 2002-737437	20020607
	R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, SI, LT, LV, FI, RO, MK, CY, AL, TR				

PRAI US 2001-296812P P 20010607
 US 2002-77383 A 20020215
 WO 2002-US18229 W 20020607

OS MARPAT 138:1057

AB This invention provides for a novel amplification procedure for nucleic acid. The method uses a wild type or mutant RNA polymerase designed to transcribe both deoxyribonucleotides and ribonucleotides. The invention provides for **oligonucleotide primers** that comprise in the following order from 5' to 3': a phage-encoded RNA polymerase recognition sequence, a **spacer sequence** comprising a sequence of from 12 to 20 nucleotides that consists of one nucleotide or two different nucleotide types, and a target complimentary sequence which can bind a segment of a target nucleic acid. The target nucleic acid can be ssDNA or comprised of RNA. The invention further provides a kit for amplifying a target nucleic acid, containing a wild type or a mutant phage RNA polymerase competent to incorporate dNTP and rNTP simultaneously into a template nucleic acid.

RE.CNT 1 THERE ARE 1 CITED REFERENCES AVAILABLE FOR THIS RECORD

ALL CITATIONS AVAILABLE IN THE RE FORMAT

AB This invention provides for a novel amplification procedure for nucleic acid. The method uses a wild type or mutant RNA polymerase designed to transcribe both deoxyribonucleotides and ribonucleotides. The invention provides for **oligonucleotide primers** that comprise in the following order from 5' to 3': a phage-encoded RNA polymerase recognition sequence, a **spacer sequence** comprising a sequence of from 12 to 20 nucleotides that consists of one nucleotide or two different nucleotide types, and a target complimentary sequence which can bind a segment of a target nucleic acid. The target nucleic acid can be ssDNA or comprised of RNA. The invention further provides a kit for amplifying a target nucleic acid, containing a wild type or a mutant phage RNA polymerase competent to incorporate dNTP and rNTP simultaneously into a template nucleic acid.

ST nucleic acid amplification phage RNA polymerase kit
oligonucleotide primer; intermediate duplex
amplification **spacer sequence** dNTP rNTP

IT 9068-38-6, **Reverse transcriptase**
RL: BSU (Biological study, unclassified); BIOL (Biological study)
(**RNaseH**-; nucleic acid amplification utilizing intermediate duplexes)

=> dup rem 14

PROCESSING COMPLETED FOR L4

L6 3 DUP REM L4 (5 DUPLICATES REMOVED)

=> d 16 1-3 bib ab kwic

L6 ANSWER 1 OF 3 CAPLUS COPYRIGHT 2005 ACS on STN
AN 2002:946303 CAPLUS
DN 138:1057
TI Nucleic acid amplification utilizing intermediate duplexes
IN Haydock, Paul V.; U'Ren, Jack
PA Saigene Corporation, USA
SO PCT Int. Appl., 69 pp.
CODEN: PIXXD2
DT Patent
LA English
FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	WO 2002098895	A1	20021212	WO 2002-US18229	20020607
	W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZM, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM				
	RW: GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR, BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG				
	US 2003050444	A1	20030313	US 2002-77383	20020215
	EP 1404697	A1	20040407	EP 2002-737437	20020607
	R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, SI, LT, LV, FI, RO, MK, CY, AL, TR				
PRAI	US 2001-296812P	P	20010607		
	US 2002-77383	A	20020215		
	WO 2002-US18229	W	20020607		

OS MARPAT 138:1057

AB This invention provides for a novel amplification procedure for nucleic acid. The method uses a wild type or mutant RNA polymerase designed to transcribe both deoxyribonucleotides and ribonucleotides. The invention

provides for **oligonucleotide primers** that comprise in the following order from 5' to 3': a phage-encoded RNA polymerase recognition sequence, a **spacer sequence** comprising a sequence of from 12 to 20 nucleotides that consists of one nucleotide or two different nucleotide types, and a target complimentary sequence which can bind a segment of a target nucleic acid. The target nucleic acid can be ssDNA or comprised of RNA. The invention further provides a kit for amplifying a target nucleic acid, containing a wild type or a mutant phage RNA polymerase competent to incorporate dNTP and rNTP simultaneously into a template nucleic acid.

RE.CNT 1 THERE ARE 1 CITED REFERENCES AVAILABLE FOR THIS RECORD
ALL CITATIONS AVAILABLE IN THE RE FORMAT

AB This invention provides for a novel amplification procedure for nucleic acid. The method uses a wild type or mutant RNA polymerase designed to transcribe both deoxyribonucleotides and ribonucleotides. The invention provides for **oligonucleotide primers** that comprise in the following order from 5' to 3': a phage-encoded RNA polymerase recognition sequence, a **spacer sequence** comprising a sequence of from 12 to 20 nucleotides that consists of one nucleotide or two different nucleotide types, and a target complimentary sequence which can bind a segment of a target nucleic acid. The target nucleic acid can be ssDNA or comprised of RNA. The invention further provides a kit for amplifying a target nucleic acid, containing a wild type or a mutant phage RNA polymerase competent to incorporate dNTP and rNTP simultaneously into a template nucleic acid.

ST nucleic acid amplification phage RNA polymerase kit
oligonucleotide primer; intermediate duplex
amplification **spacer sequence** dNTP rNTP

IT 9068-38-6, **Reverse transcriptase**

RL: BSU (Biological study, unclassified); BIOL (Biological study)
(RNaseH-; nucleic acid amplification utilizing intermediate duplexes)

L6 ANSWER 2 OF 3 MEDLINE on STN

DUPLICATE 1

AN 94118405 MEDLINE

DN PubMed ID: 7507181

TI A specific orientation of RNA secondary structures is required for initiation of reverse transcription.

AU Aiyar A; Ge Z; Leis J

CS Department of Biochemistry, Case Western Reserve University School of Medicine, Cleveland, Ohio 44106.

NC CA38046 (NCI)

P30 CA 43703 (NCI)

SO Journal of virology, (1994 Feb) 68 (2) 611-8.

Journal code: 0113724. ISSN: 0022-538X.

CY United States

DT Journal; Article; (JOURNAL ARTICLE)

LA English

FS Priority Journals; AIDS

EM 199402

ED Entered STN: 19940312

Last Updated on STN: 19970203

Entered Medline: 19940218

AB The 5' end of avian retrovirus RNA near the **primer**-binding site (PBS) forms two secondary structures, the U5-inverted repeat (U5-IR) and the U5-leader stems, and contains a 7-nucleotide sequence that anneals to the T psi C loop of the tRNA(Trp) **primer**. Mutations that disrupt any of these base pair interactions cause defects in initiation of reverse transcription both in vivo and in vitro (D. Cobrinik, A. Aiyar, Z. Ge, M. Katzman, H. Huang, and J. Leis, J. Virol. 65:3864-3872, 1991; A. Aiyar, D. Cobrinik, Z. Ge, H.-J. Kung, and J. Leis, J. Virol. 66:2464-2472, 1992). We have now examined the effect of perturbing the non-base-paired intervening "**spacer**" sequences between these secondary-structure elements. Small deletions or insertions in these intervening sequences decreased initiation of reverse

transcription in vitro. In contrast, base substitutions, which maintain the spacing distances between the structures, had no detectable effect. Additionally, a small deletion at the 3' end of the PBS caused a significant decrease in initiation of reverse transcription whereas substitution mutations again had no effect. Together, these results indicate that **reverse transcriptase** forms a complex in which the different structural elements are maintained in a specific orientation that is required for efficient initiation of reverse transcription. Specific sequence recognition of the duplex structures by **reverse transcriptase** is also required since mosaic RNAs that combine the human immunodeficiency virus type 1 PBS with avian sequences is not efficiently utilized for reverse transcription even though the **primer** used can anneal to the substituted PBS.

AB The 5' end of avian retrovirus RNA near the **primer**-binding site (PBS) forms two secondary structures, the U5-inverted repeat (U5-IR) and the U5-leader stems, and contains a 7-nucleotide sequence that anneals to the T psi C loop of the tRNA(Trp) **primer**. Mutations that disrupt any of these base pair interactions cause defects in initiation of reverse transcription both in vivo and. . . H.-J. Kung, and J. Leis, J. Virol. 66:2464-2472, 1992). We have now examined the effect of perturbing the non-base-paired intervening "**spacer**" **sequences** between these secondary-structure elements. Small deletions or insertions in these intervening sequences decreased initiation of reverse transcription in vitro. In. . . a significant decrease in initiation of reverse transcription whereas substitution mutations again had no effect. Together, these results indicate that **reverse transcriptase** forms a complex in which the different structural elements are maintained in a specific orientation that is required for efficient initiation of reverse transcription. Specific sequence recognition of the duplex structures by **reverse transcriptase** is also required since mosaic RNAs that combine the human immunodeficiency virus type 1 PBS with avian sequences is not efficiently utilized for reverse transcription even though the **primer** used can anneal to the substituted PBS.

L6 ANSWER 3 OF 3 CAPLUS COPYRIGHT 2005 ACS on STN DUPLICATE 2
 AN 1991:36720 CAPLUS
 DN 114:36720

TI Sequence, organization and transcription of the ribosomal RNA operon and the downstream tRNA and protein genes in the archaebacterium *Thermophilum pendens*

AU Kjems, Joergen; Leffers, Henrik; Olesen, Tina; Holz, Ingelore; Garrett, Roger A.

CS Kem. Inst., Aarhus Univ., Aarhus, 8000, Den.

SO Systematic and Applied Microbiology (1990), 13(2), 117-27
 CODEN: SAMIDF; ISSN: 0723-2020

DT Journal

LA English

AB The single rRNA (rRNA) operon from the extremely thermophilic archaebacterium *T. pendens* was sequenced together with the immediate downstream tRNA genes and open reading frames on both DNA strands. The genes for 16S and 23S RNA were separated by a short **spacer sequence** and were not followed by a 5S RNA gene. Sites of initiation and termination of the rRNA transcript, and its processing sites, were localized by S1 or mung bean nuclease mapping and by **primer**-directed **reverse transcriptase** anal. Initiation occurred primarily 187 nucleotides upstream from the 16S RNA gene, after an archaebacterial promoter and this was confirmed by a guanyltransferase capping experiment. The transcript terminated inefficiently before a polypyrimidine sequence 45 nucleotides downstream from the 23S RNA gene. The 16S RNA leader sequence, the spacer region and the sequence downstream from the 23S RNA can generate extensive secondary structure, including the processing stems for the 2 rRNAs. Moreover, much of this structure is supported phylogenetically by coordinated base changes. It

is proposed that some of these double helical structures are involved in transcriptional regulation. The 16S and 23S RNA sequences were aligned with those of other organisms. Secondary structures were generated from the alignments which are characteristic of the extreme thermophiles. Moreover, phylogenetic trees were derived which placed *T. pendens* close to *Thermoproteus tenax*. The downstream tRNA genes and open reading frames each exhibited an archaeobacterial promoter-like motif and a putative primary initiation site. Incomplete termination also occurred at polypyrimidine sequences. A 919-bp sequence between the 2 tRNA genes, which are located on opposing DNA strands, was rich in polypyrimidine sequences on both strands. Transcript mapping suggested that this constitutes a major termination region.

AB The single rRNA (rRNA) operon from the extremely thermophilic archaeobacterium *T. pendens* was sequenced together with the immediate downstream tRNA genes and open reading frames on both DNA strands. The genes for 16S and 23S RNA were separated by a short **spacer sequence** and were not followed by a 5S RNA gene. Sites of initiation and termination of the rRNA transcript, and its processing sites, were localized by S1 or mung bean nuclease mapping and by **primer-directed reverse transcriptase** anal. Initiation occurred primarily 187 nucleotides upstream from the 16S RNA gene, after an archaeobacterial promoter and this was confirmed by a guanyltransferase capping experiment. The transcript terminated inefficiently before a polypyrimidine sequence 45 nucleotides downstream from the 23S RNA gene. The 16S RNA leader sequence, the spacer region and the sequence downstream from the 23S RNA can generate extensive secondary structure, including the processing stems for the 2 rRNAs. Moreover, much of this structure is supported phylogenetically by coordinated base changes. It is proposed that some of these double helical structures are involved in transcriptional regulation. The 16S and 23S RNA sequences were aligned with those of other organisms. Secondary structures were generated from the alignments which are characteristic of the extreme thermophiles. Moreover, phylogenetic trees were derived which placed *T. pendens* close to *Thermoproteus tenax*. The downstream tRNA genes and open reading frames each exhibited an archaeobacterial promoter-like motif and a putative primary initiation site. Incomplete termination also occurred at polypyrimidine sequences. A 919-bp sequence between the 2 tRNA genes, which are located on opposing DNA strands, was rich in polypyrimidine sequences on both strands. Transcript mapping suggested that this constitutes a major termination region.

=>